



US009216014B2

(12) **United States Patent**
Devellian et al.

(10) **Patent No.:** **US 9,216,014 B2**
(45) **Date of Patent:** **Dec. 22, 2015**

(54) **DEVICE WITH BIOLOGICAL TISSUE SCAFFOLD FOR PERCUTANEOUS CLOSURE OF AN INTRACARDIAC DEFECT AND METHODS THEREOF**

(71) Applicant: **W.L. Gore & Associates, Inc.**, Flagstaff, AZ (US)

(72) Inventors: **Carol A. Devellian**, Topsfield, MA (US);
Robert M. Carr, Paradise Valley, AZ (US)

(73) Assignee: **W.L. Gore & Associates, Inc.**, Flagstaff, AZ (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 291 days.

(21) Appl. No.: **13/893,270**

(22) Filed: **May 13, 2013**

(65) **Prior Publication Data**
US 2013/0253538 A1 Sep. 26, 2013

Related U.S. Application Data

(60) Division of application No. 11/705,380, filed on Feb. 12, 2007, now abandoned, which is a continuation of application No. 10/453,709, filed on Jun. 3, 2003, now abandoned.

(60) Provisional application No. 60/385,274, filed on Jun. 3, 2002.

(51) **Int. Cl.**
A61B 1/32 (2006.01)
A61B 17/00 (2006.01)
A61B 17/12 (2006.01)

(52) **U.S. Cl.**
CPC **A61B 17/0057** (2013.01); **A61B 17/12122** (2013.01); **A61B 17/12172** (2013.01);
(Continued)

(58) **Field of Classification Search**

CPC A61B 17/0057; A61B 2017/00575;
A61B 2017/00641; A61B 2017/00579; A61B
2017/00588; A61B 2017/00584; A61B
2017/00592; A61B 2017/00597; A61B
2017/00601; A61B 2017/00606

See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

2,127,903 A 8/1938 Bowen
3,562,820 A 2/1971 Braun

(Continued)

FOREIGN PATENT DOCUMENTS

EP 1013227 12/1999
EP 1046375 10/2000

(Continued)

OTHER PUBLICATIONS

Ruiz et al. "The Puncture Technique: A New Method for Transcatheter Closure of Patent Foramen Ovale." Catheterization and Cardiovascular Interventions 53, Wiley-Liss, Inc., 2001, pp. 369-372.

(Continued)

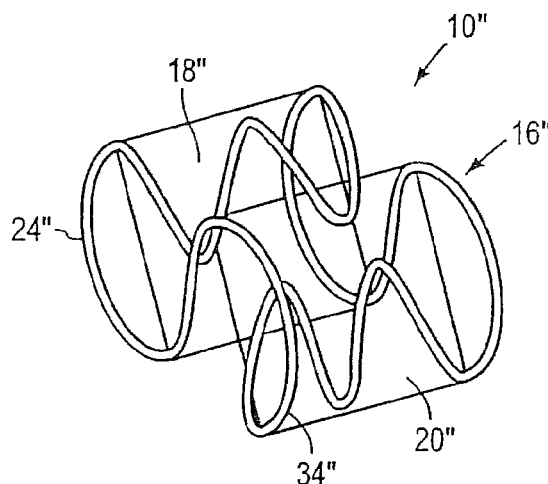
Primary Examiner — Mary Hoffman

(74) *Attorney, Agent, or Firm* — DLA Piper LLP (US)

(57) **ABSTRACT**

The invention provides an intracardiac occluder, which has biological tissue scaffolds as occlusion shells, for the percutaneous transluminal treatment of an intracardiac defect. The intracardiac occluder includes a proximal support structure supporting the proximal occlusion shell and a distal support structure supporting the distal occlusion shell. In one embodiment, biological tissue derived from the tunica submucosa layer of the porcine small intestine forms the occlusion shells.

13 Claims, 8 Drawing Sheets



(52) U.S. CL.

CPC A61B2017/00575 (2013.01); A61B
2017/00592 (2013.01); A61B 2017/00606
(2013.01); A61B 2017/1205 (2013.01); A61F
2310/00365 (2013.01)

(56)

References Cited

U.S. PATENT DOCUMENTS

3,874,388 A	4/1975	King et al.	5,904,703 A	5/1999	Gilson
3,875,648 A	4/1975	Bone	5,919,200 A	7/1999	Stambaugh et al.
3,924,631 A	12/1975	Mancusi	5,924,424 A	7/1999	Stevens et al.
4,006,747 A	2/1977	Kronenthal et al.	5,928,250 A	7/1999	Koike et al.
4,007,743 A	2/1977	Blake	5,944,738 A	8/1999	Amplatz et al.
4,425,908 A	1/1984	Simon	5,955,110 A	9/1999	Patel et al.
4,696,300 A	9/1987	Anderson	5,976,174 A	11/1999	Ruiz
4,710,192 A	12/1987	Liotta et al.	5,989,268 A	11/1999	Pugsley, Jr. et al.
4,836,204 A	6/1989	Landymore et al.	5,993,475 A	11/1999	Lin et al.
4,902,508 A	2/1990	Badylak et al.	5,993,844 A	11/1999	Abraham et al.
4,915,107 A	4/1990	Rebuffat et al.	5,997,575 A	12/1999	Whitson et al.
4,956,178 A	9/1990	Badylak et al.	6,010,517 A	1/2000	Baccaro
5,021,059 A	6/1991	Kensey et al.	6,024,756 A	2/2000	Huebsch et al.
5,037,433 A	8/1991	Wilk et al.	6,056,760 A	5/2000	Koike et al.
5,041,129 A	8/1991	Hayhurst et al.	6,077,291 A	6/2000	Das
5,108,420 A	4/1992	Marks	6,079,414 A	6/2000	Roth
5,171,259 A	12/1992	Inoue	6,080,182 A	6/2000	Shaw et al.
5,192,301 A	3/1993	Kamiya et al.	6,096,347 A	8/2000	Geddes et al.
5,222,974 A	6/1993	Kensey et al.	6,113,609 A	9/2000	Adams
5,236,440 A	8/1993	Hlavacek	6,117,159 A	9/2000	Huebsch et al.
5,257,637 A	11/1993	El Gazayerli	6,126,686 A	10/2000	Badylak et al.
5,275,826 A	1/1994	Badylak et al.	6,132,438 A	10/2000	Fleischman et al.
5,282,827 A	2/1994	Kensey et al.	6,143,037 A	11/2000	Goldstein et al.
5,284,488 A	2/1994	Sideris	6,165,183 A	12/2000	Kuehn et al.
5,304,184 A	4/1994	Hathaway et al.	6,165,204 A	12/2000	Levinson et al.
5,312,341 A	5/1994	Turi	6,171,329 B1	1/2001	Shaw et al.
5,312,435 A	5/1994	Nash et al.	6,174,322 B1	1/2001	Schneidt
5,334,217 A	8/1994	Das	6,187,039 B1	2/2001	Hiles et al.
5,354,308 A	10/1994	Simon et al.	6,190,353 B1	2/2001	Makower et al.
5,411,481 A	5/1995	Allen et al.	6,206,895 B1	3/2001	Levinson
5,413,584 A	5/1995	Schulze	6,206,907 B1	3/2001	Marino et al.
5,417,699 A	5/1995	Klein et al.	6,206,931 B1	3/2001	Cook et al.
5,425,744 A	6/1995	Fagan et al.	6,214,029 B1	4/2001	Thill et al.
5,433,727 A	7/1995	Sideris	6,217,590 B1	4/2001	Levinson
5,451,235 A	9/1995	Lock et al.	6,221,092 B1	4/2001	Koike et al.
5,460,962 A	10/1995	Kemp	6,228,097 B1	5/2001	Levinson et al.
5,478,353 A	12/1995	Yoon	6,245,080 B1	6/2001	Levinson
5,480,424 A	1/1996	Cox	6,270,515 B1	8/2001	Linden et al.
5,486,193 A	1/1996	Bourne et al.	6,277,138 B1	8/2001	Levinson et al.
5,507,811 A	4/1996	Koike et al.	6,287,317 B1	9/2001	Makower et al.
5,540,712 A	7/1996	Kleshinski et al.	6,290,674 B1	9/2001	Roue et al.
5,573,784 A	11/1996	Badylak et al.	6,299,635 B1	10/2001	Frantzen
5,601,571 A	2/1997	Moss	6,306,150 B1	10/2001	Levinson
5,618,311 A	4/1997	Gryskiewicz	6,312,446 B1	11/2001	Huebsch et al.
5,620,461 A	4/1997	Muijs Van De Moer et al.	6,315,791 B1	11/2001	Gingras et al.
5,626,599 A	5/1997	Bourne et al.	6,319,263 B1	11/2001	Levinson
5,634,936 A	6/1997	Linden et al.	6,322,548 B1	11/2001	Payne et al.
5,649,950 A	7/1997	Bourne et al.	6,334,872 B1	1/2002	Termin et al.
5,683,411 A	11/1997	Kavteladze et al.	6,342,064 B1	1/2002	Koike et al.
5,693,085 A	12/1997	Buirge et al.	6,344,049 B1	2/2002	Levinson et al.
5,702,421 A	12/1997	Schneidt	6,346,074 B1	2/2002	Roth
5,709,707 A	1/1998	Lock et al.	6,348,041 B1	2/2002	Klint
5,711,969 A	1/1998	Patel et al.	6,352,552 B1	3/2002	Levinson et al.
5,720,754 A	2/1998	Middleman et al.	6,355,052 B1	3/2002	Neuss et al.
5,725,552 A	3/1998	Kotula et al.	6,364,853 B1	4/2002	French et al.
5,733,294 A	3/1998	Forber et al.	6,375,625 B1	4/2002	French et al.
5,733,337 A	3/1998	Carr, Jr. et al.	6,375,671 B1	4/2002	Kobayashi et al.
5,741,297 A	4/1998	Simon	6,379,342 B1	4/2002	Levinson
5,776,162 A	7/1998	Kleshinski	6,379,368 B1	4/2002	Corcoran et al.
5,800,516 A	9/1998	Fine et al.	6,387,104 B1	5/2002	Pugsley, Jr. et al.
5,810,884 A	9/1998	Kim	6,398,796 B2	6/2002	Levinson
5,853,422 A	12/1998	Huebsch et al.	6,402,772 B1	6/2002	Amplatz et al.
5,855,614 A	1/1999	Stevens et al.	6,440,152 B1	8/2002	Gainor et al.
5,861,003 A	1/1999	Latson et al.	6,443,972 B1 *	9/2002	Bosma et al. 606/200
5,879,366 A	3/1999	Shaw et al.	6,460,749 B1	10/2002	Levinson et al.
5,885,619 A	3/1999	Patel et al.	6,482,224 B1	11/2002	Michler et al.
5,893,856 A	4/1999	Jacob et al.	6,488,706 B1	12/2002	Solymar
5,902,319 A	5/1999	Daley	6,494,888 B1	12/2002	Laufer et al.
			6,551,344 B2	4/2003	Thill
			6,596,013 B2	7/2003	Yang et al.
			6,623,508 B2	9/2003	Shaw et al.
			6,623,518 B2	9/2003	Thompson et al.
			6,712,836 B1	3/2004	Berg et al.
			6,726,696 B1	4/2004	Houser et al.
			8,915,958 B2 *	12/2014	Braido 623/2.11
			9,005,242 B2 *	4/2015	Cahill 606/215
			9,017,377 B2 *	4/2015	Steiner et al. 606/213
			2001/0034537 A1	10/2001	Shaw et al.
			2001/0037129 A1	11/2001	Thill

(56)

References Cited**U.S. PATENT DOCUMENTS**

2001/0044639	A1	11/2001	Levinson
2001/0049492	A1	12/2001	Frazier et al.
2002/0010481	A1	1/2002	Jayaraman
2002/0019648	A1	2/2002	Akerfeldt et al.
2002/0026208	A1	2/2002	Roe et al.
2002/0029048	A1	3/2002	Miller
2002/0032462	A1	3/2002	Houser et al.
2002/0043307	A1	4/2002	Ishida et al.
2002/0052572	A1	5/2002	Franco et al.
2002/0077555	A1	6/2002	Schwartz
2002/0096183	A1	7/2002	Stevens et al.
2002/0099389	A1	7/2002	Michler et al.
2002/0107531	A1	8/2002	Schreck et al.
2002/0111647	A1	8/2002	Khairkhahan et al.
2002/0120323	A1	8/2002	Thompson et al.
2002/0129819	A1	9/2002	Feldman et al.
2002/0169377	A1	11/2002	Khairkhahan et al.
2002/0183786	A1	12/2002	Girton
2002/0183787	A1	12/2002	Wahr et al.
2003/0028213	A1	2/2003	Thill et al.
2003/0045893	A1	3/2003	Ginn
2003/0050665	A1	3/2003	Ginn
2003/0059640	A1	3/2003	Marton et al.
2003/0065379	A1	4/2003	Babbs et al.
2003/0100920	A1	5/2003	Akin et al.
2003/0139819	A1	7/2003	DeBeer et al.
2003/0191495	A1	10/2003	Ryan et al.
2003/0195530	A1	10/2003	Thill
2004/0143291	A1	7/2004	Corcoran et al.
2004/0210301	A1	10/2004	Obermiller
2005/0043759	A1*	2/2005	Chanduszko 606/213

FOREIGN PATENT DOCUMENTS

EP	1222897	7/2002
WO	WO-95/22301	8/1995
WO	WO 96/25179	8/1996
WO	WO 96/31157	10/1996
WO	WO 97/28744	8/1997
WO	WO/9807375	2/1998
WO	WO/9918862	4/1999
WO	WO/9918864	4/1999
WO	WO/9918870	4/1999
WO	WO/9918871	4/1999
WO	WO00/27292	5/2000
WO	WO01/08600	2/2001
WO	WO01/49185	7/2001
WO	WO/0178596	10/2001
WO	WO01/93783	12/2001
WO	WO03/061481	7/2003
WO	WO03/073944	9/2003

OTHER PUBLICATIONS

International Search Report, International Application No. PCT/US03/17390, mailed on Oct. 6, 2003, 4 pgs.

SMST-2000, "Proceedings of the International Conference on Shape Memory and Superelastic Technologies," Apr. 30 to May 4, 2000, Asilomar Conference Center.

National Aeronautics and Space Administration, "55-Nitinol—The Alloy With a Memory: Its Physical Metallurgy, Properties, and Applications," NASA-SP 5110, pp. 24-25.

Kimura, et al., "Effects of Neutron Irradiation on the Transformation Behavior in Ti—Ni Alloys," Proceedings of the International Conference on Martensitic Transformations, 1992, pp. 935-940.

Ramanathan, et al., "Experimental and Computational Methods for Shape Memory Alloys," 15th ASCE Engineering Mechanics Conference, Jun. 2-5, 2002.

Shabalovskaya, "Surface, Corrosion and Biocompatibility Aspects of Nitinol as an Implant Material," Bio-Medical Materials and Engineering 12, 2002, pp. 69-109.

Uchil, "Shape Memory Alloys—Characterization Techniques," PRAMANA—Journal of Physics, vol. 58, Nos. 5 & 6, May & Jun. 2002, pp. 1131-1139.

Abraham et al., "Evaluation of the Porcine Intestinal Collagen Layer as Biomaterial" Journal of Biomed. Mater. Res., 51: 442-452 (2000).

Bailey, "The Fate of Collagen Implants in Tissue Defects," Wound Rep. Reg., 8:5-12 (2000).

Billiar et al., "Effects of Carbodiimide Crosslinking Conditions on the Physical Properties of Laminated Intestinal Submucosa," J. Biomed. Mater. Res., 56:101-108 (2001).

Edelman "Laparoscopic Herniorrhaphy with Porcine Small Intestinal Submucosa: A Preliminary Study" JSLS, 6: 203-205 (2002).

Golomb et al., "The Role of Glutaraldehyde-Induced Cross-Links in Calcification of Bovine Pericardium Used in Cardiac Valve Bioprostheses," Am. J. Pathol., 127:122-130 (1987).

Jorge-Herrero et al., "Calcification of Soft Study of Different Chemical Treatments," Tissue Employed in the Construction of Heart Valve Prostheses: Biomaterials, 12:249-252 (1991).

Huynh et al., "Remodeling of an Acellular Collagen Graft into a Physiologically Responsive Neovessel," Nature Biotechnology, 17:1083-1086 (1999).

Jux, Christian et al., "Interventional Atrial Septal Defect Closure Using a Totally Bioresorbable Occluder Matrix" Journal of the American College of Cardiology, vol. 48, No. 1, 2006: 161169.

Jux, Christian et al., "A New Biological Matrix for Septal Occlusion" Journal of Interventional Cardiology, vol. 16, No. 2, 2003:149-152.

Mullen et al., "A Prospective, Multicenter, Phase I Clinical Trial to Evaluate the Feasibility, Efficacy, and Safety of the BioSTAR Bioabsorbable Septal Repair Implant for the Closure of Atrial-Level Shunts," Circulation, 114:1962-1967 (2006).

Ramshaw et al., "Collagen-based Biomaterials" Biotechnology and Genetic Engineering Reviews, 13:335-382 (1995).

Supplemental Partial European Search Report for EP 03 75 6366 dated Jul. 23, 2008, 3 pages.

"Intestinal Collagen" Presented in Pediatric Interventional Cardiac Symposium—PICS 2007, Structural Heart Disease Symposium. Jul. 22-25, 2007, Las Vegas, Nevada. (1 page).

Schoen et al., "Long-term failure rate and morphologic correlations in porcine bioprosthetic heart valves" Am J Cardiol. Mar. 15, 1983;51(6):957-64.

* cited by examiner

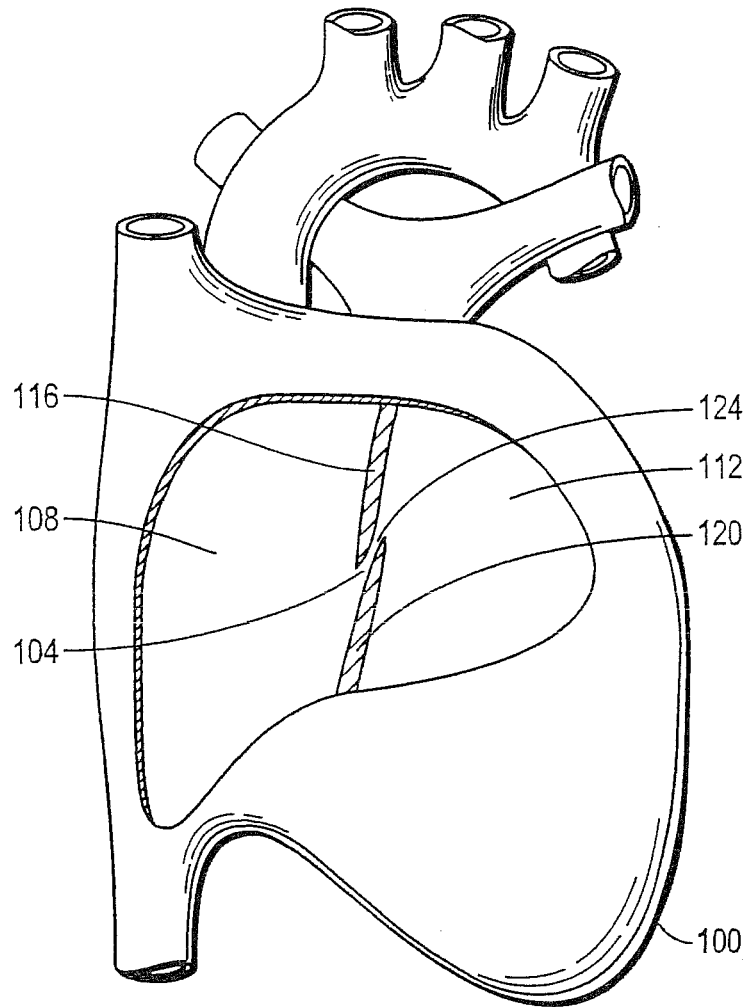


FIG. 1

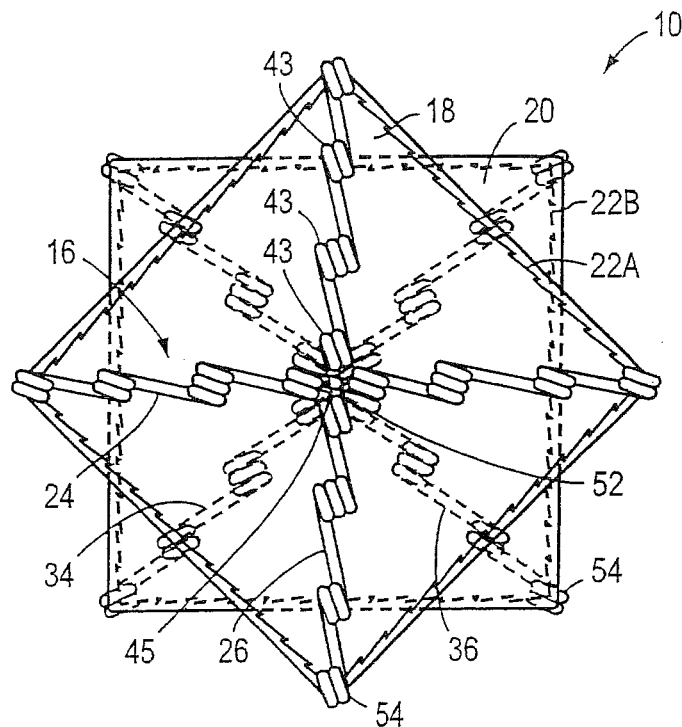


FIG. 2A

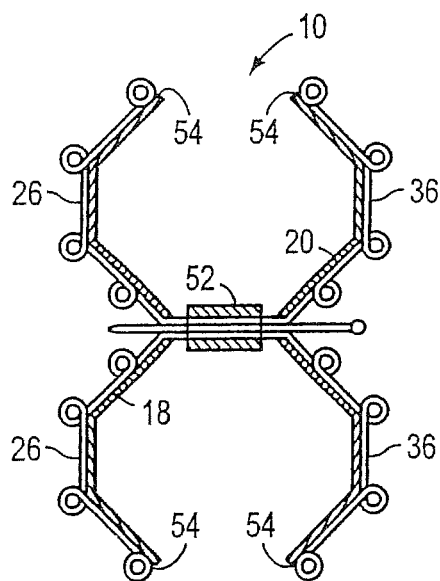


FIG. 2B

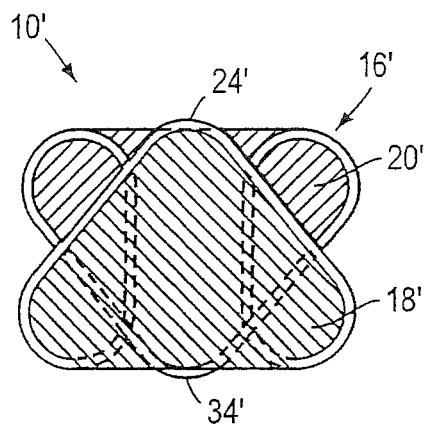


FIG. 3A

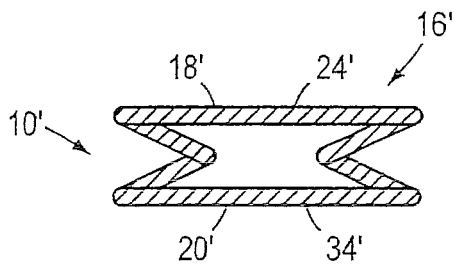


FIG. 3B

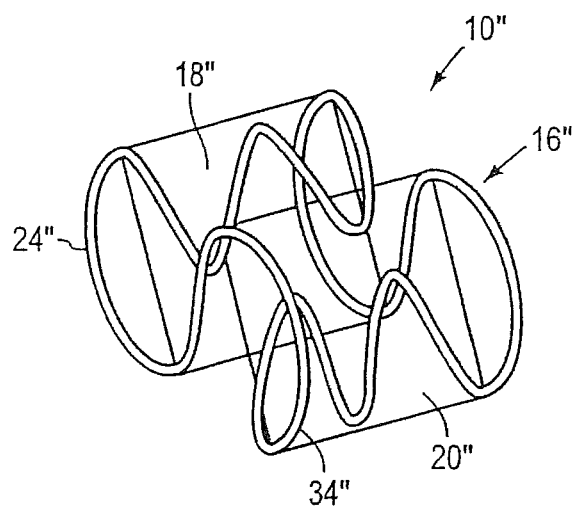


FIG. 4

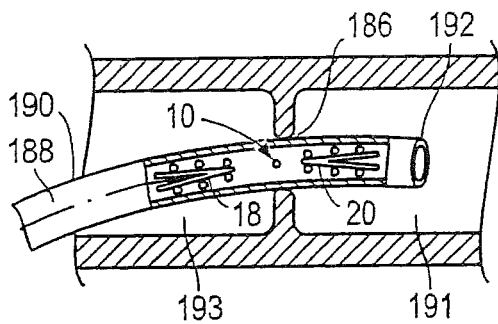


FIG. 5A

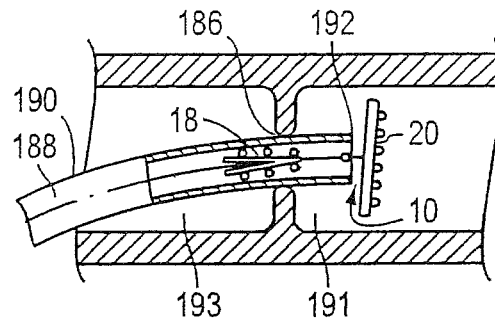


FIG. 5B

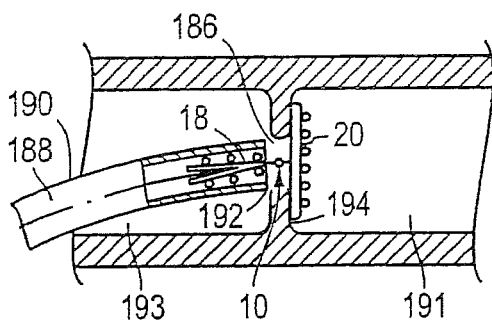


FIG. 5C

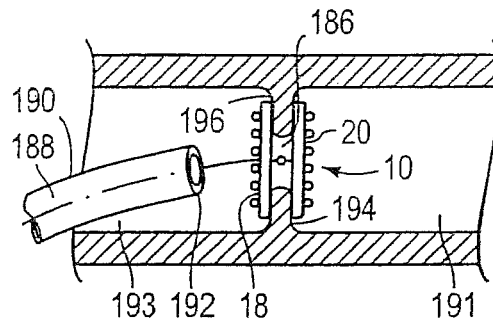


FIG. 5D

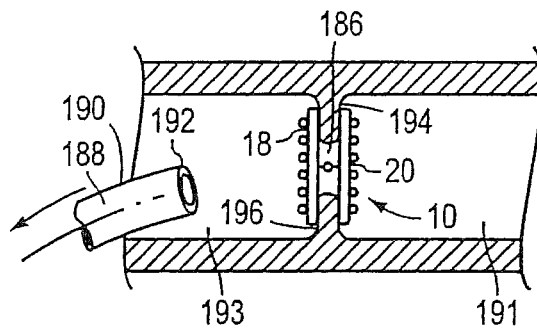


FIG. 5E

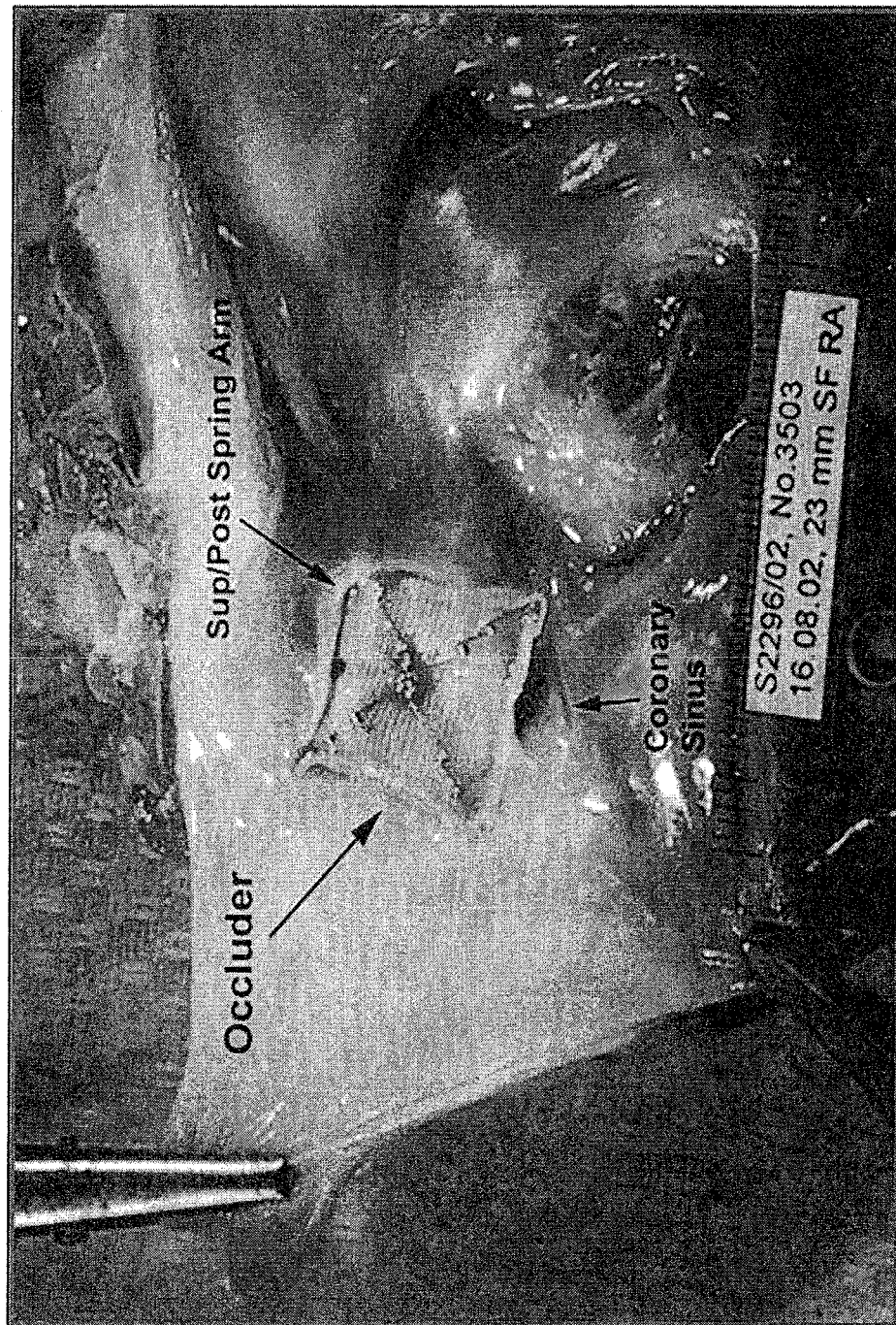


FIG. 6A

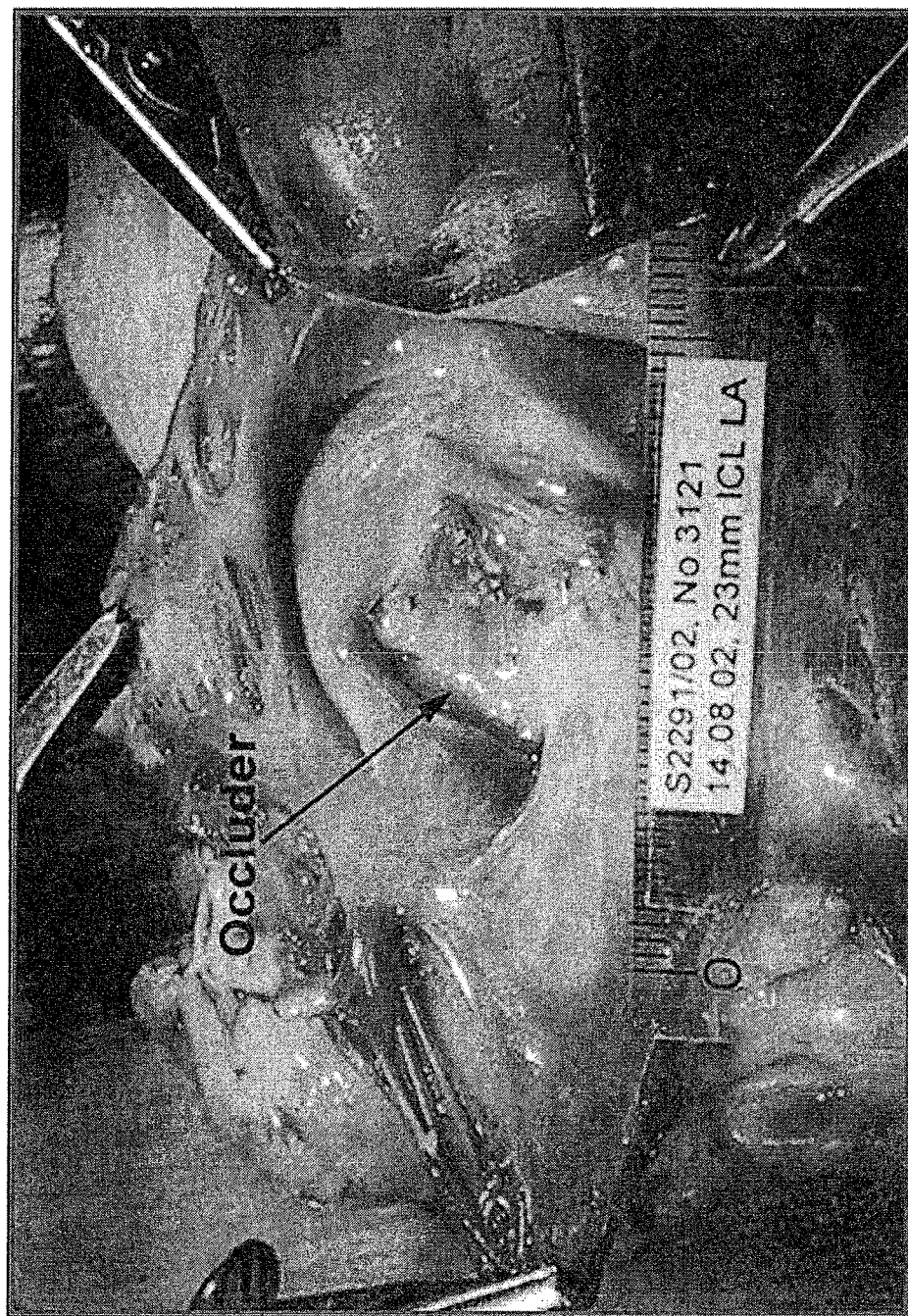


FIG. 6B



FIG. 7A



FIG. 7B

1

DEVICE WITH BIOLOGICAL TISSUE SCAFFOLD FOR PERCUTANEOUS CLOSURE OF AN INTRACARDIAC DEFECT AND METHODS THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a divisional application of U.S. application Ser. No. 11/705,380 filed Feb. 12, 2007, now pending; which is a continuation application of U.S. application Ser. No. 10/453,709 filed Jun. 3, 2003, now abandoned; which claims the benefit under 35 USC §119(e) to U.S. Application Ser. No. 60/385,274 filed Jun. 3, 2002. The disclosure of each of the prior applications is considered part of and is incorporated by reference in the disclosure of this application.

BACKGROUND OF THE INVENTION

1. Field of the Invention

The invention generally relates to devices and related methods for treating intracardiac defects. More particularly, the invention provides an intracardiac occluder with a biological tissue scaffold, and related methods, for the percutaneous closure of intracardiac defects.

2. Background Information

The human heart is divided into four compartments or chambers. The left and right atria are located in the upper portion of the heart and the left and right ventricles are located in the lower portion of the heart. The left and right atria are separated from each other by a muscular wall, the intraatrial septum, while the ventricles are separated by the intraventricular septum.

Either congenitally or by acquisition, abnormal openings, holes, or shunts can occur between the chambers of the heart or the great vessels, causing blood to flow therethrough. Such deformities are usually congenital and originate during fetal life when the heart forms from a folded tube into a four chambered, two unit system. The deformities result from the incomplete formation of the septum, or muscular wall, between the chambers of the heart and can cause significant problems. Ultimately, the deformities add strain on the heart, which may result in heart failure if they are not corrected.

One such deformity or defect, a patent foramen ovale, is a persistent, one-way, usually flap-like opening in the wall between the right atrium and left atrium of the heart. Since left atrial pressure is normally higher than right atrial pressure, the flap typically stays closed. Under certain conditions, however, right atrial pressure exceeds left atrial pressure, creating the possibility for right to left shunting that can allow blood clots to enter the systemic circulation. This is particularly worrisome to patients who are prone to forming venous thrombus, such as those with deep vein thrombosis or clotting abnormalities.

Nonsurgical (i.e., percutaneous) closure of patent foramen ovals, as well as similar intracardiac defects such as atrial septal defects, ventricular septal defects, and left atrial appendages, is possible using a variety of mechanical closure devices. These devices, which allow patients to avoid the potential side effects often associated with standard anticoagulation therapies, typically consist of a metallic structural framework that is combined with a synthetic scaffold material. The synthetic scaffold material encourages ingrowth and encapsulation of the device. Current devices typically utilize a polyester fabric, expanded polytetrafluoroethylene (ePTFE), Ivalon®, or a metal mesh as the synthetic scaffold

2

material. Such devices suffer, however, from several disadvantages, including thrombus formation, chronic inflammation, and residual leaks.

SUMMARY OF THE INVENTION

The present invention provides a device for occluding intracardiac defects. The device includes a biological tissue scaffold, as opposed to a synthetic scaffold (e.g., a polyester fabric, ePTFE, Ivalon®, or a metal mesh) as presently used by devices known in the art. In a preferred embodiment, the biological tissue scaffold is fabricated from collagen. In one embodiment, a specific type of biological tissue, derived from the tunica submucosa layer of the porcine small intestine, forms the tissue scaffold. As a result of this structure, the aforementioned disadvantages associated with the devices known in the art are minimized or eliminated.

In one aspect, the invention provides an intracardiac occluder for percutaneous transluminal treatment of an intracardiac defect. The intracardiac occluder includes a proximal support structure supporting a proximal occlusion shell and a distal support structure supporting a distal occlusion shell. The distal support structure is coupled to the proximal support structure and at least one of the occlusion shells includes a biological tissue scaffold.

Various embodiments of this aspect of the invention include the following features. The biological tissue scaffold may be a purified bioengineered type 1 collagen that may be derived from a tunica submucosa layer of a porcine small intestine. Further, in one embodiment, at least one of the support structures includes a corrosion resistant metal. Alternatively, at least one of the support structures includes a bioresorbable polymer or a biodegradable polymer. In yet another embodiment, the proximal support structure includes a plurality of outwardly extending proximal arms and the distal support structure includes a plurality of outwardly extending distal arms.

In another aspect, the invention provides a method for percutaneous transluminal treatment of an intracardiac defect in a patient. The method includes providing an intracardiac occluder as described above, positioning the intracardiac occluder proximate the intracardiac defect, and engaging the intracardiac defect with the intracardiac occluder to substantially occlude the intracardiac defect.

In one embodiment of this aspect of the invention, the intracardiac defect is engaged by positioning the proximal occlusion shell and the distal occlusion shell on different sides of the intracardiac defect. The intracardiac defect may be, for example, a patent foramen ovale, an atrial septal defect, a ventricular septal defect, or a left atrial appendage.

In yet another aspect, the invention provides a method for making an intracardiac occluder for the percutaneous transluminal treatment of an intracardiac defect. The method includes providing an overall support structure and first and second biological tissue scaffolds. The overall support structure includes a proximal support structure and a distal support structure. The method further includes coupling the first biological tissue scaffold to the proximal support structure and coupling the second biological tissue scaffold to the distal support structure. In various embodiments of this aspect of the invention, the biological tissue scaffolds are sewn, laminated, or glued to the support structures.

The foregoing and other objects, aspects, features, and advantages of the invention will become more apparent from the following description and from the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

In the drawings, like reference characters generally refer to the same parts throughout the different views. Also, the draw-

3

ings are not necessarily to scale, emphasis instead generally being placed upon illustrating the principles of the invention.

FIG. 1 is a cutaway view of a heart illustrating an intracardiac defect.

FIG. 2A is a top plan view of an intracardiac occluder according to an illustrative embodiment of the invention.

FIG. 2B is a cross-sectional view of the illustrative intracardiac occluder of FIG. 2A.

FIG. 3A is a top plan view of an intracardiac occluder according to another illustrative embodiment of the invention.

FIG. 3B is a side view of the illustrative intracardiac occluder of FIG. 3A.

FIG. 4 is a perspective view of an intracardiac occluder according to another illustrative embodiment of the invention.

FIGS. 5A-5E illustrate the stages, according to an illustrative embodiment of the invention, for delivering an intracardiac occluder to an anatomical site in the body of a patient.

FIG. 6A illustrates the results from occluding an intracardiac defect with an intracardiac occluder known in the art, 30-days after delivery of the intracardiac occluder.

FIG. 6B illustrates the results from occluding an intracardiac defect with an intracardiac occluder according to the invention, 30-days after delivery of the intracardiac occluder.

FIG. 7A illustrates the results from occluding an intracardiac defect with an intracardiac occluder known in the art, 90-days after delivery of the intracardiac occluder.

FIG. 7B illustrates the results from occluding an intracardiac defect with an intracardiac occluder according to the invention, 90-days after delivery of the intracardiac occluder.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides an intracardiac occluder for the repair of intracardiac defects, such as, for example, a patent foramen ovale, an atrial septal defect, a ventricular septal defect, and left atrial appendages. The intracardiac occluder includes a structural framework and a biological tissue scaffold adhered thereto.

FIG. 1 depicts a cutaway view of a heart 100. The heart 100 includes a septum 104 that divides a right atrium 108 from a left atrium 112. The septum 104 includes a septum primum 116, a septum secundum 120, and an exemplary intracardiac defect 124, which is to be corrected by the intracardiac occluder of the present invention, between the septum primum 116 and the septum secundum 120. Specifically, a patent foramen ovale 124 is shown as an opening through the septum 104. The patent foramen ovale 124 provides an undesirable fluid communication between the right atrium 108 and the left atrium 112. Under certain conditions, a large patent foramen ovale 124 in the septum 104 would allow for the shunting of blood from the right atrium 108 to the left atrium 112. If the patent foramen ovale 124 is not closed or obstructed in some manner, a patient is placed at high risk for an embolic stroke.

FIG. 2A depicts an intracardiac occluder 10 according to an illustrative embodiment of the invention. As shown, the intracardiac occluder 10 includes a proximal occlusion shell 18 (i.e., an occlusion shell that is closest to an operator of the intracardiac occluder 10 (e.g., a physician)), an opposite distal occlusion shell 20, and an overall support structure 16. The overall support structure 16 includes a proximal support structure 24, for supporting the proximal occlusion shell 18, and a distal support structure 34, for supporting the distal occlusion shell 20. In one embodiment, both the proximal support structure 24 and the distal support structure 34

4

include outwardly extending arms to support each of their respective occlusion shells 18, 20. As shown in FIG. 2A, for example, the proximal support structure 24 includes four outwardly extending arms 26 and the distal support structure 34 similarly includes four outwardly extending arms 36. In one embodiment, each outwardly extending arm is resiliently biased as a result of including three or more resilient coils 43 radially spaced from a center point 45. Alternatively, other resilient support structures could be used. In one embodiment, the eight arms 26, 36 are mechanically secured together by wire 52. Alternatively, other means, such as, for example, laser welding, may be used to secure the eight arms 26, 36 together. A cross-sectional view of the intracardiac occluder 10 illustrated in FIG. 2A, showing four arms 26, 36, is depicted in FIG. 2B.

FIGS. 3A and 3B depict an intracardiac occluder 10' according to another illustrative embodiment of the invention. An overall support structure 16' forms a clip and includes a proximal support structure 24', for supporting a proximal occlusion shell 18', and a distal support structure 34', for supporting a distal occlusion shell 20'.

An intracardiac occluder 10" according to yet another illustrative embodiment of the invention is illustrated in FIG. 4. Again, an overall support structure 16" forms a clip and includes a proximal support structure 24", for supporting a proximal occlusion shell 18", and a distal support structure 34", for supporting a distal occlusion shell 20".

Alternatively, the overall support structure 16 may assume any shape or configuration to form the proximal support structure 24 and the distal support structure 34.

In one embodiment, the overall support structure 16 is fabricated from a corrosion resistant metal, such as, for example, stainless steel, nitinol, or a nickel-cobalt-chromium-molybdenum alloy (e.g., MP35N). Alternatively, in other embodiments, the overall support structure 16 is fabricated from bioresorbable or biodegradable polymers.

In accordance with the present invention, the occlusion shells 18, 20, which are attached, as described below, to the proximal support structure 24 and the distal support structure 34, respectively, are made from a biological tissue scaffold. In a preferred embodiment, the tissue scaffold is fabricated from collagen. In one embodiment, a purified (acellular) bioengineered type 1 collagen derived from the tunica submucosa layer of the porcine small intestine forms the tissue scaffold. More specifically, the tunica submucosa layer, referred to hereinafter as the Intestinal Collagen Layer ("ICL"), is separated or delaminated from the other layers of the porcine small intestine (i.e., the tunica muscularis and the tunica mucosa) by any method known in the art. For example, a Bitterling sausage casing machine is used to perform the separation. Once mechanically separated from the other layers, the ICL is, in one embodiment, chemically cleaned to remove debris and other substances, other than collagen. For example, the ICL is soaked in a buffer solution at 4 degrees Celsius without the use of any detergents, or, alternatively, in a second embodiment, it is soaked with NaOH or trypsin. Other cleaning techniques known to those skilled in the art may also be used. After cleaning, the ICL is decontaminated. Any sterilization system for use with collagen, as known in the art, may be used. For example, a dilute peracetic acid solution, gamma sterilization, or electron-beam sterilization is used to decontaminate the ICL.

Alternatively, collagenous tissue from the fascia lata, pericardium, or dura matter of pigs or other mammalian sources, such as, for example, cows or sheep, may form the tissue scaffold. Additionally, in making the occlusion shells 18, 20, two or more collagen layers may be bonded together and then

5

cross-linked to produce a biocompatible material capable of being remodeled by the host cells.

In one embodiment, the biological tissue scaffold is non-porous and prevents the passage of fluids that are intended to be retained by the implantation of the intracardiac occluder **10**. In another embodiment, heparin is ionically or covalently bonded to the biological tissue scaffold to render it non-thrombogenic. In yet other embodiments, proteins or cells are applied to the biological tissue scaffold to render it non-thrombogenic and/or accelerate the healing process. Growth factors may also be applied to the biological tissue scaffold to accelerate the healing process.

Referring again to FIG. 2A, the occlusion shells **18, 20** are, in one embodiment, generally square in shape. Alternatively, the occlusion shells **18, 20** may assume other shapes. The biological tissue scaffold forming the occlusion shells **18, 20** is strong and flexible. The occlusion shells **18, 20** therefore easily attach to the overall support structure **16** and, as explained below, withstand sheath delivery to an anatomical site in the body of a patient. In one embodiment, the occlusion shells **18, 20** are sewn, as at **22A, 22B**, with any commonly used suture material (e.g., a polyester suture) that threads through the distal ends **54** of the respective arms **26, 36** of the proximal support structure **24** and the distal support structure **34**. Alternatively, the occlusion shells **18, 20** are laminated, glued, or attached by, for example, hooks or thermal welding to the proximal support structure **24** and the distal support structure **34**. In yet another embodiment, the occlusion shells **18, 20** are laminated to the overall support structure **16** and, additionally, to one another, such that the overall support structure **16** is encapsulated entirely within the occlusion shells **18, 20**.

FIGS. 5A-5E depict the stages for delivering the intracardiac occluder **10**, according to an illustrative embodiment of the invention, percutaneously to an anatomical site in the body of a patient. Referring to FIG. 5A, a sheath **190** is first inserted into the intracardiac defect **186** as is typically performed by one skilled in the art. The intracardiac occluder **10** is then loaded into the lumen **188** of the sheath **190** and advanced throughout the lumen **188** until positioned at the distal end **192** of the sheath **190**. Referring to FIG. 5B, the distal occlusion shell **20** of the intracardiac occluder **10** is released into the distal heart chamber **191** through the distal end **192** of the sheath **190**. The distal occlusion shell **20** opens automatically and resiliency. The sheath **190** is then pulled back into the proximal heart chamber **193**, as illustrated in FIG. 5C, to seat the distal occlusion shell **20** against the distal wall surface **194** of the intracardiac defect **186**. The intracardiac defect **186** is thereby occluded from the distal side. As shown in FIG. 5D, the sheath **190** is then further withdrawn a sufficient distance to allow the proximal occlusion shell **18** to be released from the distal end **192** of the sheath **190**. The proximal occlusion shell **18** opens automatically and resiliently to lie against the proximal surface **196** of the intracardiac defect **186**, occluding the intracardiac defect **186** from the proximal side. The sheath **190** is then withdrawn from the patient's body, leaving behind the opened intracardiac occluder **10**. As shown in FIG. 5E, the occlusion shells **18, 20** are positioned on either side of the intracardiac defect **186** and the intracardiac occluder **10** is permanently implanted within the body of the patient.

FIGS. 6A-6B and 7A-7B depict comparative 30-day and 90-day results, respectively, for the percutaneous closures of interventionally created intracardiac defects in sheep. Specifically, FIGS. 6A and 7A depict the 30-day and 90-day results, respectively, when an exemplary intracardiac occluder known in the art, whose occlusion shells were fab-

6

ricated from a polyester fabric (i.e., a synthetic scaffold material), is used to occlude the intracardiac defect. FIGS. 6B and 7B depict the 30-day and 90-day results, respectively, when the intracardiac occluder **10** of the instant invention, whose occlusion shells **18, 20** were fabricated from ICL, is used to occlude the intracardiac defect.

As shown, the biological tissue scaffold of the intracardiac occluder **10** of the present invention increases the rate of tissue ingrowth and, consequently, decreases the time needed to completely close the intracardiac defect. Specifically, referring now to FIG. 7B, the intracardiac occluder **10** of the present invention is barely visible after 90-days. The surrounding tissue ingrowth nearly completely envelopes the intracardiac occluder **10**. In comparison, referring now to FIG. 7A, the exemplary intracardiac occluder known in the art is still clearly visible after the same period of time.

As also shown, the intracardiac occluder **10** of the present invention naturally adheres to, and seals completely along, the edge of the intracardiac defect in a manner that is much improved from the exemplary intracardiac occluder known in the art. Additionally, in one embodiment, the biological tissue scaffold of the intracardiac occluder **10** of the present invention is non-porous. As a result, the intracardiac occluder **10** decreases the likelihood of fluid (e.g., blood) leakage through the opening.

Further advantages to the intracardiac occluder **10** of the present invention, in comparison to known intracardiac occluders, include decreased thrombogenicity, quicker endothelialization, superior biocompatibility, minimal foreign body reaction, decreased immunological and inflammatory responses, and no fibrosis.

Variations, modifications, and other implementations of what is described herein will occur to those of ordinary skill in the art without departing from the spirit and the scope of the invention as claimed. Accordingly, the invention is to be defined not by the preceding illustrative description but instead by the spirit and scope of the following claims.

What is claimed is:

1. An intracardiac occluder clip for permanent implantation in the percutaneous transluminal space to treat an intracardiac defect so as to substantially close the intracardiac defect through host tissue endothelialization, said occluder comprising:

a resilient wire forming opposing proximal and distal support structures for occlusion shells, wherein each support structure is formed of two external stems and two internal stems forming a M-shaped arcuate length of the wire, wherein both external stems of each M-shaped support structure are joined to the external stems of the opposing M-shaped support structure;

a proximal occlusion shell providing a scaffold for endothelialization, wherein the shell is formed of a non-porous bioresorbable sheet and is fitted securely between the external and internal stems of the M-shaped proximal support structure; and,

a distal occlusion shell providing a scaffold for endothelialization, wherein the shell is formed of a non-porous bioresorbable sheet and is fitted securely between the external and internal stems of the M-shaped distal support structure.

2. The occluder of claim 1, wherein the bioresorbable sheet is purified bioengineered type 1 collagen.

3. The occluder of claim 2, wherein the purified bioengineered type 1 collagen is derived from a tunica submucosa layer of the intestine.

4. The occluder of claim 1, wherein the wire is fabricated from a corrosion resistant metal.

7

5. The occluder of claim 1, wherein at the wire is fabricated a bioresorbable polymer.

6. The occluder of claim 1, wherein the wire is fabricated from a biodegradable polymer.

7. The occluder of claim 1, wherein the collagen sheet is treated to be non-thrombogenic. 5

8. A method for treating a septal defect in a heart, the method comprising:

(a) compressing the occluder of claim 1 into the distal end of a catheter sheath;

(b) introducing the sheath into the heart and through the septal defect to position its distal end on the distal surface of the septal defect; 10

(c) extending the distal occlusion shell out of the sheath to automatically and resiliently seat the shell against the distal side of the septal defect; 15

(d) withdrawing the sheath to position its distal end of the sheath at the proximal surface of the septal defect;

8

(e) extending the proximal occlusion shell out of the sheath to automatically and resiliently seat the shell against the proximal side of the septal defect; and,

(f) releasing the occluder to leave it in the septal defect and withdrawing the sheath from the heart.

9. The method of claim 8, wherein the intracardiac defect is a patent foramen ovale.

10. The method of claim 8, wherein the intracardiac defect is an atrial septal defect.

11. The method of claim 8, wherein the intracardiac defect is a ventricular septal defect.

12. The method of claim 8, wherein the intracardiac defect is a left atrial appendage.

13. The method of claim 8, further comprising step (g), confirming after at least 90 days that endothelialization of the occlusion shells has occurred.

* * * * *